

Acknowledgment. We take pleasure in acknowledging the generous support we have received from CIBA-GEIGY, Ltd., and the continuing interest and encouragement of Professor Albert Wettstein. Warm thanks are due to Dr. Hans Fritz and his colleagues (Spectroscopic services, CIBA-GEIGY, Ltd.) for the recording and discussion of numerous spectra.

R. B. Woodward,* J. Gosteli, I. Ernest
R. J. Friary, G. Nestler, H. Raman
R. Sitrin, Ch. Suter, J. K. Whitesell

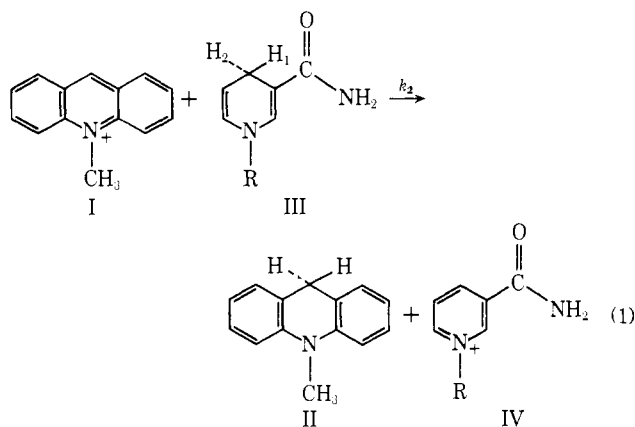
Woodward Research Institute
CH-4002 Basel, Switzerland

Received July 23, 1973

Model Dehydrogenase Reactions. Reduction of *N*-Methylacridinium Ion by Reduced Nicotinamide Adenine Dinucleotide and Its Derivatives

Sir:

Despite the central importance of NADH and NADPH in biochemical oxidation-reductions, relatively few facile nonenzymic reductions by 1,4-dihydronicotinamides are known. "Model" reactions of this type are of potential value in that they may provide helpful clues as to the mechanism of action of NAD⁺- and NADP⁺-dependent dehydrogenases.¹⁻⁹ In the present communication we wish to report a new nonenzymic reaction of dihydronicotinamides. Specifically, we have found that *N*-methylacridinium ion (I) is rapidly reduced to *N*-methylacridan (II) by β -NADH and a variety of dihydronicotinamide derivatives at room temperature in essentially quantitative yield (eq 1). Since the reverse reaction has not been observed



experimentally, the reaction must be strongly thermodynamically favored in the direction written. The re-

(1) D. Mauzerall and F. H. Westheimer, *J. Amer. Chem. Soc.*, **77**, 2261 (1955).

(2) R. H. Abeles, R. F. Hutton, and F. H. Westheimer, *J. Amer. Chem. Soc.*, **79**, 712 (1957).

(3) C. H. Suetter and D. B. Metzler, *Biochim. Biophys. Acta*, **44**, 23 (1960).

(4) D. C. Dittmer and R. A. Fouty, *J. Amer. Chem. Soc.*, **86**, 91 (1964).

(5) K. A. Schellenberg, G. W. McLean, H. C. Lipton, and P. S. Lietman, *J. Amer. Chem. Soc.*, **89**, 1948 (1967).

(6) (a) T. P. Goldstein, Abstracts, 152nd National Meeting of the American Chemical Society, New York, N. Y., Sept 11-16, 1966, Abstract C196; (b) J. J. Steffens and D. M. Chipman, *J. Amer. Chem. Soc.*, **93**, 6694 (1971).

(7) D. J. Creighton and D. S. Sigman, *J. Amer. Chem. Soc.*, **93**, 6314 (1971).

(8) M. Brüstlein and T. C. Bruice, *J. Amer. Chem. Soc.*, **94**, 6548 (1972).

(9) S. Shinkai and T. C. Bruice, *J. Amer. Chem. Soc.*, **94**, 8258 (1972).

duction of *N*-methylacridinium (I) is considerably more rapid than other nonenzymic transhydrogenation reactions such as the hydrogen exchange between NAD⁺ and NADH¹⁰ and the reduction of *N*-benzyl-3-acetylpyridinium chloride¹¹ and the acetyl analog of NAD⁺ by NADH.^{12,13}

The second-order rate constants for the reduction of I by a series of dihydronicotinamides are summarized in Table I. Since II and the various oxidized nicotin-

Table I. Rates of Reduction of *N*-Methylacridinium Ion by a Series of Dihydronicotinamides^a

Compound	R	k_2 , $M^{-1} \text{ sec}^{-1}$
β -NADH (IIIa)	ADPR	101.2 ± 2.4^b
β -NADH (IIIa)	ADPR	98.2 ± 5.1^c
β -NMNH (IIIb)	Ribose 5-phosphate	41.9 ± 1.6^b
IIIc'	$\text{CH}_2\text{CH}_2\text{CH}_2$	2040 ± 50^d
IIIc''	$\text{CH}_2\text{CH}_2\text{CH}_2$; $\text{H}_1 = \text{D}$	1620 ± 50^d
IIIc'''	$\text{CH}_2\text{CH}_2\text{CH}_2$; $\text{H}_1 = \text{H}_2 = \text{D}$	1398 ± 60^e

^a In these experiments, the concentration of reactants did not exceed $10^{-4} M$. Under these conditions, the reaction was strictly first order with respect to each component. ^b pH 8.0, 0.1 *M* phosphate buffer; 25°. ^c pH 7.0, 0.1 *M* phosphate buffer; 25°. ^d pH 8.4, 0.01 *M* phosphate buffer; 25°. ^e pH 8.7, 0.01 *M* phosphate buffer; 25°.

amides do not absorb strongly above 320 nm, the reaction can be conveniently assayed by following either (a) the disappearance of absorption at 358 nm where I absorbs very intensely ($\epsilon = 2.6 \times 10^4 M^{-1} \text{ cm}^{-1}$) and the dihydronicotinamides absorb with variable intensities depending on the nature of R; (b) the disappearance of the characteristic absorbance of I in the region of 420 nm; or (c) the disappearance of the intense fluorescence of I at 490 nm. The latter two methods for assaying the reaction are particularly useful when the dihydronicotinamides are present in large excess relative to I.

The production of II and IV was confirmed by nuclear magnetic resonance and mass spectra, ultraviolet and visible absorption spectra, and thin-layer chromatographic analysis of the products isolated from the reaction mixture. Independently prepared samples of II and the various nicotinamides were used as internal standards in these procedures. Spectral analyses of the reaction mixtures prior to isolation of the products were completely consistent with the stoichiometry indicated in eq 1.

The reaction proceeds by direct hydrogen transfer and is unaffected by reagents known to affect the rates of free-radical reactions. Direct hydrogen transfer was demonstrated in two ways. First, I was reduced with NADH in D₂O and the deuterium content of the resulting *N*-methylacridan was determined from its appearance potential mass spectrum. The intensity of the P + 1 peak (*m/e* 196) for this sample and that for II generated from the oxidation of NADH in H₂O were equal and corresponded to the intensity (15.72%) predicted from the natural isotopic abundance for a parent

(10) J. Ludoweig and A. Levy, *Biochemistry*, **3**, 373 (1964).

(11) G. Cilento, *Arch. Biochem. Biophys.*, **88**, 352 (1960).

(12) M. J. Spiegel and G. P. Drysdale, *J. Biol. Chem.*, **235**, 2498 (1960).

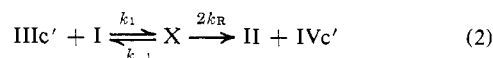
(13) G. R. Drysdale, M. J. Spiegel, and P. Strittmatter, *J. Biol. Chem.*, **236**, 2323 (1961).

ion with a composition of $C_{14}H_{13}N$.¹⁴ As a further check that no solvent protons or deuterons were incorporated in II and that the reaction proceeds by direct hydrogen transfer, 4-R-NADH- d_1 and IIIc'' were used as reductants for I in H_2O . In each case, the mass spectrum of II isolated from the reaction mixture possessed a more intense P + 1 peak than expected on the basis of the normal isotopic abundance. The free-radical quenching agents, dihydroquinone and 4-*tert*-butylcatechol, do not inhibit the rate of the reduction of I by NADH. When I and NADH are present at concentrations of 4.77×10^{-5} and 2.77×10^{-5} M, respectively, neither dihydroquinone nor 4-*tert*-butylcatechol at concentrations of 1×10^{-3} M has any significant effect on the second-order rate constant for the reaction. The demonstration of direct hydrogen transfer as well as the insensitivity of the rate to the presence of free-radical quenching agents indicate the reaction proceeds *via* a mechanism formally similar to a hydride transfer mechanism.

Examination of the isotope effects for the reduction of I by the various isotopic forms of *N*-propyldihydronicotinamide (IIIc', IIIc'', and IIIc''') suggests that the reduction of I is not a simple bimolecular process. *N*-Propyldihydronicotinamide instead of NADH was used as the reductant to study isotope effects since the conformation of the coenzyme renders the (*pro-R*)- and (*pro-S*)-hydrogens of the dihydronicotinamide ring chemically nonequivalent.^{15,16} Hence, interpretation of the isotope effects would be more complex with NADH than with *N*-propyldihydronicotinamide. If the reduction of I by *N*-propyldihydronicotinamide proceeds by a bimolecular mechanism without the formation of any kinetically significant intermediate, the ratio of undeuterated II (*m/e* 195) to deuterated II (*m/e* 196) obtained after reduction of I with IIIc'' should approximate the primary kinetic isotope effects determined for the reduction of I by IIIc'' and IIIc''' when secondary isotope effects are assumed to be one. Since the isotopic partitioning ratio is 5.4 ± 1.0 ¹⁷ and the primary kinetic isotope effects obtained by comparing the rates of reduction by IIIc'' and IIIc''' to IIIc' (Table I) are 1.70 ± 0.28 and 1.46 ± 0.10 , respectively, a bimolecular mechanism is inconsistent with the experimental data assuming no significant secondary isotope effects.¹⁵ The only way the kinetic isotope

effects for IIIc'' can be made to correspond to the isotope partitioning ratio is if the secondary isotope effect for hydrogen transfer is 0.74 ± 0.06 .¹⁹ Since secondary isotope effects for reactions which involve conversion of a carbon atom from an sp^3 to an sp^2 hybridization are usually greater than one, a simple bimolecular reaction mechanism is inconsistent with the observed isotope effects.

The divergence of the isotope effects measured kinetically and from product analysis demands the existence of at least one kinetically important intermediate during the course of the reaction whose rate of formation is partially rate limiting. The simplest kinetic scheme possible in this case is indicated in eq 2



where X designates the intermediate whose precise nature cannot be deduced from the data presently available. Steffens and Chipman^{6b} have reported isotope effects for the reduction of trifluoroacetophenone by IIIc' and IIIc'' similar to those reported here for the reduction of I. They have proposed a kinetic scheme similar to that indicated in eq 2 and have suggested that the intermediate, X, is a noncovalent complex, possibly of a charge-transfer nature, which forms prior to hydrogen transfer. Although this type of an intermediate is consistent with the observed isotope effects, the value of k_1 that can be estimated for the formation of the complex composed of III and either I or trifluoroacetophenone would be of the order of magnitude of $10^3 M^{-1} sec^{-1}$ or $10^{-3} M^{-1} sec^{-1}$, respectively. Since these rate constants are substantially lower than the rates of formation of stacked dimers of aromatic compounds such as proflavine, where the rate of dimerization is $7.9 \times 10^8 M^{-1} sec^{-1}$,²⁰ either the charge transfer or noncovalent nature of these compounds can be questioned or more than one noncovalent complex exists on the reaction pathway. Some support for charge-transfer intermediates comes from recent kinetic studies on the reduction of a series of flavine derivatives by NADH and *N*-propyldihydronicotinamide which have indicated that noncovalent complex formation prior to hydrogen transfer may take place during this reaction.²¹

Additional data will be necessary before the generality of obligatory intermediate formation in nonenzymic dihydronicotinamide reductions becomes clear. Yet for the reaction systems discussed here, and possibly for the zinc ion catalyzed reduction of 1,10-phenanthroline-2-carboxaldehyde by *N*-propyldihydro-

are experimentally determined second-order rate constants, k_H is the rate of hydrogen transfer from C-4 bound to two hydrogen atoms, and k_H/k_H is the secondary isotope effect for hydrogen transfer. For a strict bimolecular reaction, comparison of the observed second-order rate constants for the reduction by dideuterio- (IIIc''') and dihydro- (IIIc') nicotinamide yields the following simple relationship: $k_2^{IIIc'''} / k_2^{IIIc'} = k_H/k_D$, where k_D is the rate of deuterium transfer from a C-4 bound to two deuterium atoms.

(19) In order for the kinetic isotope effects for the dideuterionicotinamide (IIIc''') to be consistent with the isotope partitioning ratio, a bimolecular mechanism requires that the product, $(k_H/k_H) \cdot (k_D/k_D)$, be equal to 0.272. But since the data for the monodeuterated derivative demand that k_H/k_H be 0.74, k_H/k_H must be significantly greater than k_D/k_D . This appears to be unlikely and serves as added evidence against a simple bimolecular mechanism.

(20) D. H. Turner, G. W. Flynn, S. K. Lundberg, L. D. Faller, and N. Sutin, *Nature (London)*, **239**, 215 (1972).

(21) T. C. Bruice, L. Main, S. Smith, and P. Y. Bruice, *J. Amer. Chem. Soc.*, **93**, 7327 (1971).

(14) R. M. Silverstein and G. Clayton Bassler, "Spectrophotometric Identification of Organic Compounds," Wiley, New York, N. Y., 1968.

(15) N. J. Oppenheimer, L. J. Arnold, and N. O. Kaplan, *Proc. Nat. Acad. Sci. U. S.*, **68**, 3200 (1971).

(16) R. H. Sarma and N. O. Kaplan, *Biochemistry*, **9**, 539 (1970).

(17) The *N*-methylacridan product analyzed for deuterium was isolated from a 100-ml reaction mixture composed of 4.4×10^{-4} M I and 4.3×10^{-4} M IIIc'' which had been allowed to react for 4 hr. The reaction mixture was extracted with three 25-ml aliquots of reagent grade chloroform which were pooled and twice washed with 10-ml aliquots of water. After the chloroform was dried, the solution was evaporated to dryness and the resulting *N*-methylacridan was analyzed for deuterium by mass spectrometry. All determinations of the isotopic composition of *N*-methylacridan were performed at the appearance potential of the P - 1 (*m/e* 194) peak of undeuterated *N*-methylacridan.

(18) For a strict bimolecular reaction, the isotopic composition of II produced by reduction with IIIc'' is given by $II(m/e 195)/II(m/e 196) = k_H/k_D$, where $II(m/e 196)$ is the observed intensity minus the intensity due to the isotopic abundance of the *m/e* 195 peak, k_H is the rate of hydrogen transfer from C-4 which is bound to one hydrogen and one deuterium, and k_D is the rate of deuterium transfer from C-4 which is bound to one hydrogen atom and one deuterium atom. As pointed out by Steffens and Chipman,^{6b} a primary isotope effect for dihydronicotinamide reduction can be obtained from kinetic experiments using the monodeuterated dihydronicotinamide, IIIc'', since $k_2^{IIIc''}/k_2^{IIIc'} = 2k_H/(k_H + k_D) = (2k_H/k_H)/[1 + (k_D/k_H)]$ where $k_2^{IIIc'}$ and $k_2^{IIIc''}$

nicotinamide,²² some type of intermediate seems essential. Since *N*-propylidihydronecotinamide reduces I roughly 50,000 times faster than trifluoroacetophenone,^{6b} intermediate formation is apparently important for a wide range of reaction rates. A more complete description of the chemical nature of these intermediates should provide a better understanding of the mechanism of catalysis of NAD⁺ and NADP⁺-dependent dehydrogenases.

Acknowledgments. This research was supported by U. S. Public Health Service Grant No. AM-12789. We wish to thank Professors G. J. Popják and E. L. Smith for helpful discussions and Dr. Popják for access to the mass spectrometer in his laboratory.

(22) D. J. Creighton, Ph.D. Thesis, University of California at Los Angeles (1972).

(23) Supported by Public Health Service Training Grant No. GM 00364.

(24) Alfred P. Sloan Research Fellow, 1972-1974.

Donald J. Creighton,²³ Joseph Hajdu
Gregory Mooser, David S. Sigman*²⁴

Department of Biological Chemistry
University of California at Los Angeles, School of Medicine
Los Angeles, California 90024

Received June 1, 1973

Large Polar Effects in the Oxidation of Hexadecanoic (Palmitic) Acid by Nitric Acid

Sir:

Free radicals are noted for their insensitivity to polar effects. For example, photochlorination of 1-chlorobutane with Cl₂ at 68° gave equal rates of attack on C₂ and C₃,¹ and photochlorination of octanoic acid with Cl₂ or *t*-BuOCl in CCl₄ gave comparable amounts of attack on C₄-C₇.² This subject has been treated in reviews.³⁻⁵ The major exceptions are reactions involving nitrogen cation (aminium) radicals.^{6,7} These exhibit large polar effects. Typical is the 80% selectivity for ω-1 photochlorination in C₆-C₈ acids⁷ and esters⁶ and the >90% ω-1 selectivity found in C₆-C₈ alcohols.^{7,8}

Another type of free radical reaction has now been found which shows large polar effects. This is the nitric acid oxidation of hexadecanoic acid, Table I. At low conversion (6%), 77% of the diacids are C₁₀-C₁₅ showing a high selectivity for attack at positions remote from the carboxyl group. As the oxidation progresses, the longer diacids cleave in the middle to produce two molecules of shorter diacids so that the distribution of diacids shifts toward C₄-C₈ (Table I) and obscures the initial high selectivity for remote attack.

The selectivity for remote attack accounts for the facts that little CO₂ or acetic acid is produced and that the net

(1) C. Walling and M. F. Mayahi, *J. Amer. Chem. Soc.*, **81**, 1485 (1959).

(2) N. Deno, R. Fishbein, and C. Pierson, *ibid.*, **92**, 1451 (1970).

(3) G. A. Russell, "Free Radicals," Vol. I, J. K. Kochi, Ed., Wiley, New York, N. Y., 1973, p 275.

(4) E. S. Huyser, "Free-Radical Chain Reactions," Wiley-Interscience, New York, N. Y., 1970.

(5) M. L. Poutsma, "Methods in Free-Radical Chemistry," Vol. 1, E. S. Huyser, Ed., Marcel Dekker, New York, N. Y., 1969, p 79.

(6) F. Minisci, *Synthesis*, **1** (1973).

(7) N. Deno, "Methods in Free-Radical Chemistry," Vol. 3, E. S. Huyser, Ed., Marcel Dekker, New York, N. Y., 1972, p 135.

(8) N. Deno, W. E. Billups, R. Fishbein, C. Pierson, R. Whalen, and J. C. Wyckoff, *J. Amer. Chem. Soc.*, **93**, 438 (1971).

Table I. Relative Yields of Dicarboxylic Acids from the Oxidation of 2.56 g of Hexadecanoic Acid with 30 ml of 70% HNO₃ at 90°

No. of carbons in dicarboxylic acid	Relative yields			
	4 hr		24 hr	120 hr
	EGSS-X	SE-30	EGSS-X	EGSS-X
4	0	0	3	8
5	3	0	4	16
6	4	0	7	24
7	4	1	12	26
8	(5) ^a	4	20	16
9	7	8	20	8
10	13	11	14	2
11	13	14	9	0
12	15	17	6	0
13	16	(17) ^a	3	0
14	11	17	2	0
15	9	(11) ^b	0	0
16	0	0	0	0

^a The value is a mean of the preceding and following value. Direct measurement was prevented because of overlap with unidentified band (total band area 31). Value was estimated from data on EGSS-X column.

weight of isolated acids increases as the reaction progresses, Table I. It also accounts for the fact that the rate of disappearance of dibasic acids increases with chain length on treatment with 62% HNO₃.⁹ The rate constants (in min⁻¹) were <10⁵ for C₄-C₆ and 2 × 10⁻⁴, 7 × 10⁻⁴, 3 × 10⁻³, and 2 × 10⁻² for C₇-C₁₀. The data in Table I also show that the longer chain diacids are selectively oxidized.

The increase in weight in going from reactant to products makes this an attractive method for the production of C₅-C₈ diacids. On the basis that the distribution at 6% conversion indicates the initial cleavage and given the distribution at 120 hr, the theoretical yield of diacids is 3.0 g. This is in good agreement with the 2.9 g isolated and further shows the absence of oxidations other than the remote oxidation pattern described.

For the early stages (6% conversion), there was concern that keto acids were present in the products (hydroxy acids were unlikely because hydroxy compounds instantly produce copious NO₂ on contact with 70% HNO₃). The general agreement between gc analyses on the polar EGSS-X and the nonpolar SE-30 columns (Table I) indicated that keto esters were not a major problem. In agreement, an infrared spectrum showed a keto band at 1410 cm⁻¹ that was only about 5% of the area of the ester carbonyl band at 1440 cm⁻¹. However, a band which is suspected of being due to a keto ester appeared between C₉ and C₁₀ on the EGSS-X column and coincided with the C₁₅ band on the SE-30 column. A similar pattern was found in model studies on the oxidation of 12-hydroxystearic acid with 70% HNO₃ at 90°.

The preliminary results are reported now because of (1) the industrial importance of oxidizing fatty acids to long chain diacids and (2) the unusually large polar effects found for this free radical reaction.

There is no direct evidence identifying the attacking radical in the HNO₃ oxidation, although the abundance

(9) G. Gut, R. V. Falkenstein, and A. Guyer, *Helv. Chim. Acta*, **49**, 481 (1966).